APPENDIX D:

STEM CELL TABLES

COMPENDIUM OF SCIENTIFIC PUBLICATIONS REGARDING THE ISOLATION AND CHARACTERIZATION OF STEM CELLS

The following tables provide an overview of information about stem cells that have been derived from mice and humans. The tables summarize published research that characterizes cells that are capable of developing into cells of multiple germ layers (i.e., multipotent or pluripotent) or that can generate the differentiated cell types of another tissue (i.e., plasticity) such as a bone marrow cell becoming a neuronal cell. The tables do not include information about cells

considered progenitor ore precursor cells or those that can proliferate without the demonstrated ability to generate cell types of other tissues.

The tables list the tissue from which the cells were derived, the types of cells that developed, the conditions under which differentiation occurred, the methods by which the cells were characterized, and the primary references for the information.

O Tissue	rigin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Bone marrow	Hematopoietic stem cell (HSC)	Cardiac muscle	Cardiac injury induced in mice Injected labeled HSCs into injured heart	Measurement of green fluorescent protein (GFP) in regenerating cardiac cells Measurement of cardiac-specific protein and gene expression Cardiac-function tests	Orlic et al., 2001
	HSC	Epithelial cells of the liver, skin, lung, esophagus, stomach, small and large intestine	Transplantation of HSCs into lethally irradiated female mice	Detection of antibodies to cellular and cell- surface proteins Cell staining Probing for Y chromo- some-positive cells	Krause et al., 2001
	HSC	Cholangiocyte Hepatocyte	Purification of HSCs from bone marrow Transplantation of HSCs into mice with liver- enzyme deficiency	Observation of normalized liver function and regenerating hepatocytes Measurement of expression of hematopoietic and hepatic cell-surface proteins	Lagasse et al., 2000
	HSC	Platelet Red blood cell White blood cell	Hematopoietic growth factors: interleukin-3, interleukin-6, granulocyte-colony stimulating factor, erythropoietin, and thrombopoietin	Detection of antibodies to cell-surface proteins Colony-forming assays Immunophenotyping	Spangrude et al., 1991 Morrison et al., 1995
	HSC Side population (SP)	Skeletal muscle	Lethal irradiation of female mice Induced muscle injury Purified bone marrow transplanted into mice	Measurement of dystrophin expression in regenerating muscle cells Fluorescence-activated cell sorting (FACS) Probing for Y chromo- some-positive cells	Gussoni et al., 1999
	Mesenchymal stem cell (MSC)	Adipocyte Chondrocyte Osteoblast Tenocyte	Dexamethasone Vitamin D₃ Bone morphogenetic protein-2 (BMP-2)	Detection of antibody binding to cell-surface proteins Immunofluorescence	Friedenstein et al., 1976 Pereira et al., 1995 Prockop, 1997

Ori Tissue	gin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Bone marrow (cont.)	MSC	Astrocyte Neuron	Injection of MSC into brain of immunocompromised neonatal mice	Detection of cell-surface markers by using antibodies and immunofluorescence	Kopen et al., 1999
	MSC	Astrocyte Neuron	Epidermal growth factor Brain-derived neurotrophic factor B-mercaptoethanol Retinoic acid	Immunofluorescence Cell sorting	Sanchez-Ramos et al., 2000
	MSC	Neuron	Stromal cells expanded as undifferentiated cells B-mercaptoethanol Butylated hydroxyanisole	Detection of numerous neuron-specific proteins via staining	Woodbury et al., 2000
	MSC	Skeletal muscle	5-azacytidine and amphotericin B	Observation of myotubes Staining for myocytes	Wakitani et al., 1995
	MSC and/or HSC	Astrocyte Microglia Oligodendrocyte	Induced injury to neural tissue Bone marrow transplantation	Detection of antibodies to cell-surface proteins	Eglitis and Mezey, 1997
	MSC and/or HSC	Cardiac muscle	Bone marrow transplantation of 5-azacytidinetreated cells into mice with induced cardiac muscle injury	Cell staining for cardiac muscle proteins Measurement of blood pressure Electron microscopy Observation of beating cells in vitro Measurement of atrial natriuretic peptide Staining cells for muscle proteins	Tomita et al., 1999 Makino et al., 1999
	MSC and/or HSC	Hepatocyte	Suppression of liver cell proliferation Induced injury to liver Bone marrow transplantation	Staining cells Antibody labeling of cell- surface markers	Taniguchi et al., 1996 Petersen et al., 1999 Theise et al., 2000

Ori Tissue	gin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Bone marrow (cont.)	MSC and/or HSC	Neuron	Induced neural tissue injury Bone marrow transplantation into female mice	Detection of antibodies to cell-surface proteins Probing for Y chromosome-containing neurons	Mezey et al., 2000 Brazelton et al., 2000
	MSC, HSC, or side population (SP)	Cardiac muscle Skeletal muscle	Lethal irradiation of mice Bone marrow transplantation from normal male donor mice into mice with induced muscle degeneration	Probing for Y chromosome-containing muscle cells Detection of expression of myoregulatory proteins	Bittner et al., 1999
	MSC, HSC, or SP	Skeletal muscle	Induced muscle tissue injury Transplantation of genetically marked bone marrow into immunodeficient mice	Histologic observation of muscle regeneration Detection of antibodies to cell-surface proteins Myogenic differentiation factor transcript expression	Ferrari et al., 1998
	SP	Cardiomyocyte Vascular endothelia	Transplanted into lethally irradiated mice with ischemic damage to cardiac tissue	Immunohistochemistry Staining for cardiomyocte marker (alpha-actin) and endothelial marker (flt-1)	Jackson et al., 2001
Brain	Neural stem cell (NSC)	Astrocyte Neuron Oligodendrocyte	Basic fibroblast growth factor Epidermal growth factor	Detection of antibodies to neural cell-specific proteins	Reynolds et al., 1996 Doetsch et al., 1999 Johansson et al., 1999
	NSC	Red blood cell White blood cell	Transplantation of NSC into irradiated mice	Flow cytometry analysis Genetic labeling assay Detection of antibodies to cell surface proteins	Bjornson et al., 1999

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Brain (cont.)	NSC	Skeletal muscle	Transplantation of NSCs into mice In vitro co-culture with myogenic cells	Observation of differentiated skeletal muscle cells Analysis of muscle cell-specific proteins and gene expression	Galli et al., 2000
Embryo- blastocyst inner-cell mass	Embryonic stem (ES)	Adipocyte	Retinoic acid Insulin, T3 (thyroid hormone), and Leukemia inhibitory factor (LIF)	Observation of adipocyte differentiation Measurement of adipocyte enzyme activity Measurement of adipocyte-specific gene expression	Dani et al., 1997
	ES	Astrocyte Glial precursor Oligodendrocyte	Cells cultured in neurogenic medium with basic fibroblast growth factor Epidermal growth factor Platelet-derived growth factor Transplanted glial precursor cells into myelin-deficient mice	Observation of spinal cord remyelination Electron microscopy Antibodies to neural cell- specific proteins	Brustle et al., 1999
	ES	Astrocyte Midbrain neuron Neural precursor Neuron Oligodendrocyte	Retinoic acid Cell selection through transgene conferring drug resistance Co-culture with stromal cells	Examination of cell morphology and neuron- specific markers Cell-specific markers Detection of dopamine production	Bain et al., 1995 Strubing et al., 1995 Li et al., 1998 Lee et al., 2000 Kawasaki et al., 2000
	ES	Astrocyte Neuron Oligodendro- cyte	Retinoic acid	Observation of functional synapses Measurement of neurotransmitters	Slager, et al., 1993 Gottlieb, et al., 1999
	ES	Astrocyte Oligodendrocyte	Retinoic acid Fetal calf serum (10%) B-mercaptoethanol	Antibodies to neural cell- specific proteins Cytochemistry	Fraichard et al., 1995

Ori Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Embryo- blastocyst inner-cell mass (cont.)	ES ES	Cardiac muscle Skeletal muscle Smooth muscle	Retinoic acid Dimethyl sulfoxide Transplantation of muscle cells into mice	Histology Detection of cell-specific proteins Cytochemistry	Dinsmore et al., 1996
	ES	Cardiomyocyte	LIF, retinoic acid Fibroblast feeder cells	Histology and observation of beating cardiomyocyte Detection of specific cardiac cell-gene expression and cardiomyocyte surface proteins	Doetschman et al., 1985 Maltsev et al., 1993 Wobus et al., 1995
	ES	Cardiomyocyte	Cell selection through genetic labeling of ES Injection of ES into mouse heart	Detection of genetically labeled cardiomyocytes Electrophysiological studies	Bader et al., 2000
	ES	Cardiomyocyte	LIF Purification of cardiomyocytes from ES culture by genetic labeling and selection	Observation of functional cardiomyocyte grafts in heart Immunohistology	Klug et al., 1996
	ES	Cardiomyocyte	Culture of ES with LIF Selection of cardiomyocytes through genetic labeling Injection of cardiomyocytes into mouse heart	Microscopy and cell- receptor studies Observation of cardiomyocyte differentiation and contractility Analysis of cardiomyocyte gene expression	Westfall et al., 1997
	ES	Chondrocyte (cartilage-forming cell)	BMP-2 and BMP-4	Staining of mature chondrocytes Measurement of chondrocyte-specific gene expression and proteins	Kramer et al., 2000

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Embryo- blastocyst inner-cell mass (cont.)	ES	Dendritic (immune cell)	Culture on stromal cell layer Interleukin-3 Granulocyte- macrophage stimulating factor	Immune-function assays Immunophenotyping	Fairchild et al., 2000
	ES	Embryoid bodies (EBs) consisting of structures that contain tissues of the three embryonic germ layers: endoderm, mesoderm, and ectoderm Teratocarcinoma	ES cultured in suspension without feeder cell layer Absence of LIF Injection of ESs into mice	Observation of differentiation into multiple tissue types of the germ layers of blood, skeletal and cardiac muscle, primitive gastrointestinal and neural tissue Growth of tumor containing tissues from embryonic germ layer	Evans and Kaufman, 1981
	ES	ES self-renewal	LIF Culture on feeder cell layer	Observation of extensive ES proliferation and self- renewal	Evans and Kaufman, 1981
	ES	Endothelial	Culture on collagen substrate Hematopoietic growth factors Semisolid media EB implanted peritoneal cavity	Observation of capillary formation	Risau et al., 1988
	ES	Endothelial Smooth muscle Vascular progenitor	Culture over collagen-IV matrix Absence of LIF Vascular endothelial growth factor	Electron microscopy: observation of endothelial and smooth muscle vascular structures Detection of endothelial cell marker by immunochemistry Detection of smooth muscle markers by immunochemistry	Yamashita et al., 2000

Ori Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Embryo- blastocyst inner-cell mass (cont.)	ES	HSC and erythroid	Interleukin-6 Absence of LIF and cell feeder layer Culture on collagen substrate Hematopoietic growth factors Semisolid media BMP-4	Antibodies against surface markers FACS Immunophenotyping	Wiles and Keller, 1991 Johansson and Wiles, 1995 Perkins et al., 1998
	ES	Keratinocyte (skin)	B-mercaptoethanol Implantation of ES cells in mice	Microscopy Immunofluorescence Observation of skin tissue differentiation Measurement of keratin	Bagutti et al., 1996
	ES	Lymphoid precursor Lymphocyte	Culture of ES in low oxygen concentration (5%) without hematopoietic growth factors	Antibodies to lymphoid cell-surface proteins Analysis of antibody production and lymphocyte receptors	Potocnik et al., 1994
	ES	Macrophage	Interleukin-3 and macrophage colony stimulating factor	Immunophenotyping Immune-function assays	Lieschke and Dunn, 1995
	ES	Mast	Lethal mutations in ES cells Culture of EBs in media containing interleukin-3, stem cell factor	Transplantation of cells into mast cell-deficient mice Immunologic- and inflammation-function tests Analysis of gene expression	Johansson and Wiles, 1995 Tsai et al., 2000
	ES	Melanocyte	Dexamethasone Stromal cell layer Steel factor	Morphology studies Reactivity to growth factors Expression of melanogenic markers	Yamane et al., 1999

Ori Tissue	gin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Embryo- blastocyst inner-cell mass (cont.)	ES	Neuron	Expression of noggin cDNA in ES Expression of neuronal determination gene EB exposed to retinoic acid	Detection of antibodies to neuronal proteins	O'Shea, 1999
	ES	Oligodendrocyte	Retinoic acid Induced spinal cord injury Transplantation of ES- derived cells into spinal cord of mice	Detection of remyelination in spinal cord Antibodies to oligodendrocyte-specific proteins	Liu et al., 2000
	ES	Osteoblast (bone cell)	Co-cultured with fetal mouse osteoblasts Dexamethasone, retinoic acid, ascorbic acid, B-glycerophosphate	Microscopy; observation of mineralized bone nodules Histochemistry	Buttery et al., 2001
	ES	Pancreatic	Insertion of insulin-gene promoter into ES	Antibodies to cellular proteins Measurement of insulin, glucagon, somatostatin Observation of islet-like organization of cells Transplantation of cells into diabetic mice with resultant lowering of blood glucose	Soria et al., 2000
	ES	Pancreatic islet-like	Serum-free media Absence of feeder-cell layer Basic fibroblast growth factor Nicotinamide	Detection of antibodies to cellular and cell- surface proteins	Lumelsky et al., 2001
	ES	Skeletal muscle	Overexpression of insulin- like growth factor-2 in ES through gene insertion Dimethyl sulfoxide	Observation of myocyte differentiation Measurement of myocyte-specific gene expression and proteins	Prelle et al., 2000

Ori Tissue	gin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Embryo- blastocyst inner-cell mass (cont.)	ES	Skeletal muscle	Transforming growth factor-beta and retinoic acid ES co-culture with stromal cells Fetal calf serum B-mercaptoethanol	Observation of myocyte differentiation Detection of functional muscle cell receptors Measurement of myocyte-specific gene expression	Slager et al., 1993 Rohwedel et al., 1994
	ES	Smooth muscle	Retinoic acid and db-cAMP Culture over collagen IV matrix Vascular endothelial growth factor Platelet-derived growth factor-BB	Electron microscopy observation of vascular structures Detection of smooth muscle markers: SMA, CGA7	Drab et al., 1997 Yamashita et al., 2000
	ES	Smooth muscle	Platelet-derived growth factor	FACS Detection of smooth muscle cell proteins	Hirashima et al., 1999
	ES	White blood cell	Interleukin-3 Transplantation of ESs into lymphocyte- deficient mice	Measurement of lymphocyte-specific gene expression Radioimmunoassay	Wiles and Keller, 1991
	ES	White blood cell	Transplantation of ES cells into lymphocyte-deficient mice	Histology Immunophenotyping Antibodies to cell- specific proteins	Rathjen et al., 1998
Gonadal ridge (fetal)	Embryonic primordial germ cell	Endoderm Mesoderm Ectoderm	"Reprogramming" primordial germ cells: culture of primordial germ cell with LIF, basic fibroblast growth factor and Steel factor	Histology Immunocytochemistry	Matsui et al., 1992

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Liver	HSC	HSC All blood cell lineages	Enrichment of cell populations through immunoselection Purification of CD45+ liver cells Selection of cells with HSC markers Transplantation of HSCs into lethally irradiated mice	Colony-forming assays Detection of in vitro growth of hematopoietic colonies by flow cytometry and cell sorting Liver-derived cells reconstituted from bone marrow of transplanted mice FACS	Taniguchi et al., 199
Pancreas	Pancreatic ductal epithelial cell	Alpha, beta, and delta pancreatic islet	Stem cells isolated from prediabetic adult, nonobese mice Cells cultured for an extensive period Pancreatic cells transplanted into diabetic mice	Analysis of pancreatic cell gene expression and differentiation markers Glucose challenge test in vitro Observation of reversal of insulin-dependent diabetes in mice with transplants	Ramiya et al., 2000
	Unselected pancreatic cells	Hepatocyte	Pancreatic cells transplanted into mice with liver-enzyme deficiency	Detection of normalized liver function in mice Histological evidence of donor-derived hepatocytes	Wang et al., 2001
Skeletal muscle	Muscle	Adipocyte	Long-chain fatty acids Thiazolidinediones	Assays of adipocyte enzyme function Observation of adipocyte differentiation Detection of adipocyte-specific gene expression	Grimaldi et al., 1997
	Muscle	Osteoclast and osteocyte Osteoprogenitor	Exposure of donor cells to BMP-2 Retroviral transfection of cells with vector and transplantation into severe combined immunodeficient mice (SCID)	Detection of ectopic bone formation Detection of musclederived cells Co-localization with osteocalcin-producing cells in newly formed bone matrix	Bosch et al., 2000

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Skeletal muscle (cont.)	Muscle Satellite	HSC Myocyte precursor	Isolation of transcription factor Pax7 as a gene expressed specifically in satellite cell-derived myoblasts	Detection of Pax7 ⁺⁻ and Pax7 ⁺ muscle cells in hematopoietic and myogenic cells	Seale et al., 2000
	Muscle Satellite or SP	All blood cell lineages HSC	Transplant of muscle- derived cells into lethally irradiated mice	Observation of engraftment of muscle cells in bone marrow Antibodies to hematopoietic cell markers FACS	Jackson et al., 1999 Gussoni et al., 1999
	Satellite	Myocyte Myocyte precursor	Induced tissue injury; mechanical and denervation stress Transcription factor expression	Detection of myocyte progenitor and myocyte- specific proteins and mRNA transcripts	Megeney et al., 199
Spinal cord	NSC	Astrocyte Neuron Oligodendrocyte	Basic fibroblast growth factor Epidermal growth factor	Detection of antibodies to neural cell proteins	Weiss et al., 1996

Appendix D.ii. Published Reports on Isolation and Differentiation of Human Fetal Tissue Germ Cells

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Gonadal ridge	Primordial germ cell	Embryoid bodies	SDEC line of embryoid body derived cells transplanted into rats paralyzed with a virus induced motor neuron degeneration	Functional assessment of rat locomotion and righting ability (turning from supine to prone) Histopathologic examination of motor axons Immunohistochemistry of mature neurons: NeuN+ and 68-kilodalton neurofilament	Kerr et al., 2001
	Primordial germ cell	Embryoid bodies with neural cells, vascular endothelium, muscle cells, endodermal derivatives	Leukemia inhibitory factor, Basic fibroblast growth factor	Clonal expression, polymerase chain reaction Ethidium bromide fluorescence detection Surface markers: 68-kilodalton neurofilament, neuron-specific enolase, tau, vimentin, human nestin, galactocerebroside, O4, SMI32	Shamblott et al., 2001
	Primordial germ cell	Embryoid bodies with three germ layers: endoderm, mesoderm, ectoderm	Leukemia inhibitory factor, Basic fibroblast growth factor	Detection of surface markers: SSEA-1, SSEA-3, SSEA-4, TRA-1-60, TRA-1- 81	Shamblott et al., 1998

Appendix D.iii. Published Reports on Isolation and Differentiation of Human Embryonic Stem Cells

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Human embryo (from in vitro fertilization (IVF))	Blastocyst inner- cell mass	Ectoderm Endoderm Mesoderm Neuronal progenitor cell	Leukemia inhibitory factor Injection into severe combined immunodeficient (SCID) mice	Developed two lines (HES-1, HES-2) Clonal expression Polymerase chain reaction Surface markers: SSEA-1, SSEA-4, TRA-1-60, GTCM-2	Reubinoff et al., 2000
	Blastocyst innercell mass (H9 clone line from Thomson et al., 1998)	Cardiomyocyte	Embryoid body formation (See Itskovitz-Eldor et al., 2000)	Visualization of contracting areas in embryoid bodies Immunohistochemistry for cardiac myosin heavy chain, alpha-actinin, desmin, cardiac troponin I, and antinaturetic protein.	Assady et al., 2001
	Blastocyst innercell mass (H9 clone line from Thomson et al., 1998)	Cardiomyocyte	Embryoid body formation	Polymerase chain reaction for cardiac-specific genes and transcription factors	Kehat et al., 2001
	Blastocyst innercell mass (H9 clone line from Thomson et al., 1998)	Cardiomyocyte Endoderm Hematopoietic Neuron	Leukemia inhibitory factor Basic fibroblast growth factor Collagenase or trypsin/EDTA to induce embryoid body	Clonal expression Polymerase chain reaction Surface markers: gamma-globin, 68- kilodalton neurofilament, alpha-fetoprotein, albumin	ltskovitz-Eldor et al., 2000
	Blastocyst innercell mass (H9 clone line from Thomson et al., 1998)	Ectoderm: brain, skin, adrenal Endoderm: liver, pancreas Mesoderm: muscle, bone, kidney, urogenital, heart, hematopoietic, hematopoietic	Basic fibroblast growth factor, transforming growth factor beta 1, activin-A, bone morphogenic protein 4 hepatocyte growth factor, epidermal growth factor, beta nerve growth factor, retinoic acid	Clonal expression Polymerase chain reaction Surface markers	Schuldiner et al., 2000

Ori Tissue	gin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Human embryo (from in vitro fertilization (IVF) (cont.)	Blastocyst inner- cell mass (H9 clone line from Thomson et al., 1998)	Ectoderm: neural epithelium, embryonic ganglia, stratified squamous epithelium Endoderm: gut epithelium Mesoderm: cartilage, bone, smooth muscle, striated muscle	Injection of cell lines into severe combined immunodeficient mice Leukemia inhibitory factor Type IV collagenase	Surface markers: SSEA-3, SSEA-4, TRA-160, TRA-181, alkaline phosphatase Radioimmunoassay detection: alpha- fetoprotein and human chorionic gonadotropin	Thomson et al., 1998
	Blastocyst innercell mass (H9 clone line from Thomson et al., 1998)	Pancreatic beta cell	Embryoid body formation (See Itskovitz-Eldor et al., 2000) No leukemia inhibitory factor or basic fibroblast growth factor	Immunohistochemistry for insulin Polymerase chain reaction for insulin, IPF1/PDX1, Ngn3, beta- actin, Glut-1, Glut-2, glucokinase, and Oct 4	Assady et al., 2001

EC

Neuron

Appendix D.iv. Published Reports on Isolation and Differentiation of Human Embryonic Carcinoma Stem Cells					
O: Tissue	rigin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Terato- carci- noma	Embryonic carcinoma (EC)	Endodermal progenitor cell	Absence of feeder cell layer Bone morphogenetic protein-2 Retinoic acid	Analysis of stem cell marker-gene transcription Immunochemistry Immunofluorescence	Roach et al., 1994 Pera and Herszfeld, 1998
	EC	Neuron	EC transplanted into mouse brain	Observation of functional synapses Immunochemistry	Trojanowski et al., 1993
	EC	Glial Neuron	Retinoic acid	Measurement of mRNA for GABA(A) receptor-chloride complex Recording of whole-cell voltage-clamp measurements in differentiated cells in the presence of GABA(A) receptor antagonists and activators (bicuculline and flurazepam, respectively)	Reynolds et al., 1994
	EC	Glial Neuron	Retinoic acid	Detection of neurons with HNK-1 antibody Measurement of acetylcholine synthesis and detection of high- affinity uptake sites for GABA	McBurney et al., 1988

Retinoic acid

Morphology and histology

Analysis of neuronspecific proteins Andrews, 1984

Ori	igin	Cell Types	Differentiation	Methods of	Reference
Tissue	Cell Type	Developed	Conditions	Characterization	
Terato- carci- noma or teratoma	EC	Tumors containing tissue types from endoderm, mesoderm, and ectoderm	Bone morphogenetic protein-7 EC cells cultured without feeder cell layer Transplantation of EC cells into mice	Morphology, histology, and cell staining Observation of tissue types from endoderm, mesoderm, and ectoderm Observation of extended self-renewal of EC cells Analysis of chromosomes and specific genes Detection of cell-specific proteins Cytochemical assay	Andrews et al., 1984 Thompson et al., 1984 Pera, 1989

Appendix D.v. Published Reports on Isolation and Differentiation of Human Adult Stem Cells						
O Tissue	rigin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference	
Blood	Circulatory Skeletal	Adipocyte Osteocyte	Leukemia inhibitory factor (LIF) Transplantation of stem cells into bg-nu-xid immunocompromised mice	Antibody labeling Polymerase chain reaction	Kuznetsov, 2001	
Bone marrow	Angioblast (endothelial precursor)	Mature endothelia and newly formed blood vessels	Angioblasts isolated by mobilizing peripheral blood with granulocytecolony stimulating factor Angioblasts injected into rats with experimental myocardial infarction	Observation of neovascularization within myocardium from transplanted cells Detection of improved cardiac function in experimental animals	Kocher et al., 2001	
	Hematopoietic stem cell (HSC)	Hepatocyte Cholangiocyte	Bone marrow transplantation	Probed for presence and function of Y chromosome-containing liver cells Measured expression of liver-specific proteins Immunochemistry	Alison et al., 2000 Theise et al., 2000	
	Human marrow stromal	Stromal-derived cell engrafted in rat brain	Isolation of marrow stromal cell from human volunteers; injection of stromal cell into rat brain	Observation of engraffment, migration, and survival of stromalderived cell in rat brain Observation of loss of stromal cell functions Antibodies to cell-surface proteins	Azizi et al., 1998	
	Mesenchymal stem cell (MSC)	Adipocyte Chondrocyte Osteocyte	Fetal bovine serum, dexamethasone, isobutylxanthine, insulin, ascorbate, indomethacin, transforming growth factor-B3, and glycerol phosphate	Histology and immunofluorescence Detection of lipids and specific enzyme activity of adipocytes and osteocytes Specific staining for chondrocytes	Pittenger et al., 1999	

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Bone marrow (cont.)	MSC	Neuron	Prolonged expansion of MSCs as undifferentiated cells β-mercaptoethanol (BME) Butylated hydroxyanisole (BHA)	Histology Detection of numerous neuron-specific proteins via staining and antibody binding	Woodbury et al., 2000
	MSC	Neuron	MSCs cultured with fetal rat brain cells Epidermal growth factor Brain-derived neurotrophic factor	Detection of nestin and nestin-gene expression Detection of neuron- specific proteins	Sanchez-Ramos et al., 2000
	MSC	Adipocyte Bone marrow stromal cell Cardiomyocyte Chondrocyte Myocyte Thymic stromal cell	MSCs isolated from bone marrow Transplantation of MSCs into fetal sheep	Analysis of human gene expression in sheep tissues Confirmed presence of human cells by immunohistochemistry	Liechty et al., 2000
Bone marrow (fetal)	HSC	HSC Red blood cell lineages White blood cell lineages	Enrichment of hematopoietic cell populations by cell selection Transplantation of bone marrow and thymus cells into mice	Establishment of long- term multilineage cultures of hematopoietic colonies Fluorescence-activated cell sorting (FACS) Engraftment of hematopoietic cells in mice	Baum et al., 1992
Brain	Neural stem cell (NSC)	Muscle cell	Exposure of NSCs to myoblasts Dissociation of NSC clusters Transplantation of human NSCs into mice with induced muscle injury	Observation of differentiated skeletal muscle cells from primary and culturederived NSCs Demonstration of NSC engraftment in mice by detection of expression of specific genes	Galli et al., 2000

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Brain (adult and neonatal)	Neural progenitor cell (NPC)	Astrocyte Neuron Oligodendrocyte	NPCs cultured in medium containing glutamine, amphotericin-B, antibiotics, fetal calf serum, basic fibroblast growth factor, epidermal growth factor, and platelet-derived growth factor AB Transplantation of human central nervous system stem cells (hCNS-SCs) into mice	Observation of functional engraftment of NPCs into mouse brain Antibody labeling of neuronal cell-surface proteins	Palmer et al., 2001
Brain (fetal)	Human central nervous system stem cell (hCNS-SC)	Astrocyte Neuron Oligodendrocyte	Fibroblast growth factor- 2, epidermal growth factor, lymphocyte inhibitory factor, neural survival factor-1, brain- derived and glial-derived neurotrophic factors	Observation of neurosphere formation and self-renewal of hCNS-SCs Demonstration of engraftment, proliferation, migration, and neural differentiation of hCNS-SCs FACS	Uchida et al., 2000
Fat	Stromal vascular cell fraction of processed lipoaspirate	Adipocyte precursor Osteocyte precursor Chondrocyte precursor Myocyte precursor	Co-cultured with mouse adipocytes, isobutylmethylxanthine, dexamethasone Co-cultured with human osteoblasts, insulin, indomethacin, antibiotic/antimycotic dexamethasone, ascorbate, b-glycerophosphate, antibiotic/antimycotic Co-cultured with human skeletal myocytes, insulin, transforming growth factor-B, ascorbate, antibiotic/antimycotic dexamethasone, hydrocortisone, antibiotic/antimycotic	Staining for lipid accumulation Staining for alkaline phosphatase activity Staining for bone formation Staining for proteoglycan-rich matrix Antibody binding to collagen II Visualization of multinucleation Staining for muscle protein: myosin Antibody binding to MyoD1	Zuk et al., 2001

Or Tissue	igin Cell Type	Cell Types Developed	Differentiation Conditions	Methods of Characterization	Reference
Liver (fetal)	HSC	Hematopoietic progenitor cell (HPC) Red blood cell lineages White blood cell lineages	Co-culture of HSCs with mouse stromal cells Implantation of fetal hematopoietic liver cells into immunocom- promised mice	Demonstration of differentiation into red and white blood cell lineages through colony- forming assays and detection of surface markers characteristic of the hematopoietic system	McCune et al., 1988 Namikawa et al., 1990
Pancreas	Nestin-positive islet-derived progenitor cell (NIP)	Pancreatic Hepatic	NIPs obtained from pancreatic islets and cultured for extended periods	Observation of extended proliferative, self-renewing, and multipotent capacity Expression of hepatic and exocrine pancreatic markers Demonstration of ductal and endocrine pancreatic features Production of insulin and glucagons	Zulewski et al., 2000
Umbilical cord blood	HPC	Most red and white blood cell lineages	Collection and sorting Stimulation with colony- stimulating factors and interleukin-3	Demonstration of multipotent progenitor, granulocyte-macrophage, and erythroid cell lines	Broxmeyer et al., 1989
	HSC Mesenchymal progenitor cell (MPC)	Most red and white blood cell lineages Osteoblasts Adipocytes	Mixtures of dexamethasone, ß-glycerol, ascorbate, insulin, isobutyl-methylxanthine, and indomethacin	Cell morphology Cytochemical analysis of osteoblast and adipocyte products Immunophenotyping	Erices et al., 1999

Appendix D.vi.

REFERENCES

- Alison, M.R., Poulsom, R., Jeffery, R., Dhillon, A.P., Quaglia, A., Jacob, J., Novelli, M., Prentice, G., Williamson, J., and Wright, N.A. (2000). Hepatocytes from non-hepatic adult stem cells. Nature. 406, 257.
- Andrews, P.W., Damjanov, I., Simon, D., Banting, G.S., Carlin, C., Dracopoli, N.C., and Fogh, J. (1984). Pluripotent embryonal carcinoma clones derived from the human teratocarcinoma cell line Tera-2. Differentiation in vivo and in vitro. Lab. Invest. 50, 147-162.
- Assady, S., Maor, G., Amit, M., Itskovitz-Eldor, J., Skorecki, K.L., and Tzukerman, M. (2001). Insulin production by human embryonic stem cells. Diabetes, 50, http://www.diabetes.org/Diabetes_Rapids/Suheir_Assady_ 06282001.pdf.
- Azizi, S.A., Stokes, D., Augelli, B.J., DiGirolamo, C., and Prockop, D.J. (1998). Engraftment and migration of human bone marrow stromal cells implanted in the brains of albino rats—similarities to astrocyte grafts. Proc. Natl. Acad. Sci. U. S. A. 95, 3908-3913.
- Bader, A., Al Dubai, H., and Weitzer, G. (2000). Leukemia inhibitory factor modulates cardiogenesis in embryoid bodies in opposite fashions. Circ. Res. 86, 787-794.
- Bagutti, C., Wobus, A.M., Fassler, R., and Watt, F.M. (1996).
 Differentiation of embryonal stem cells into keratinocytes:
 comparison of wild-type and β(1) integrin-deficient cells.
 Dev. Biol. 179, 184-196.
- Bain, G., Kitchens, D., Yao, M., Huettner, J.E., and Gottlieb, D.I. (1995). Embryonic stem cells express neuronal properties in vitro. Dev. Biol. 168, 342-357.
- Baum, C.M., Weissman, I.L., Tsukamoto, A.S., Buckle, A.M., and Peault, B. (1992). Isolation of a candidate human hematopoietic stem-cell population. Proc. Natl. Acad. Sci. U. S. A. 89, 2804-2808.
- Bittner, R.E., Schofer, C., Weipoltshammer, K., Ivanova, S., Streubel, B., Hauser, E., Freilinger, M., Hoger, H., Elbe-Burger, A., and Wachtler, F. (1999). Recruitment of bone-marrowderived cells by skeletal and cardiac muscle in adult dystrophic mdx mice. Anat. Embryol. (Berl) 199, 391-396.
- Bjornson, C.R., Rietze, R.L., Reynolds, B.A., Magli, M.C., and Vescovi, A.L. (1999). Turning brain into blood: a hematopoietic fate adopted by adult neural stem cells in vivo. Science. 283, 534-537.
- Bosch, P., Musgrave, D.S., Lee, J.Y., Cummins, J., Shuler, F., Ghivizzani, S.C., Evans, C., Robbins, P.D., and Huard, J. (2000). Osteoprogenitor cells within skeletal muscle. J. Orthop. Res. 18, 933-944.
- Brazelton, T.R., Rossi, F.M., Keshet, G.I., and Blau, H.M. (2000). From marrow to brain: expression of neuronal phenotypes in adult mice. Science. 290, 1775-1779.

- Broxmeyer, H.E., Douglas, G.W., Hangoc, G., Cooper, S., Bard, J., English, D., Arny, M., Thomas, L., and Boyse, E.A. (1989). Human umbilical cord blood as a potential source of transplantable hematopoietic stem/progenitor cells. Proc. Natl. Acad. Sci. U. S. A. 86, 3828-3832.
- Brustle, O., Jones, K.N., Learish, R.D., Karram, K., Choudhary, K., Wiestler, O.D., Duncan, I.D., and McKay, R.D. (1999).
 Embryonic stem cell-derived glial precursors: a source of myelinating transplants. Science. 285, 754-756.
- Buttery, L.D., Bourne, S., Xynos, J.D., Wood, H., Hughes, F.J., Hughes, S.P., Episkopou, V., and Polak, J.M. (2001).
 Differentiation of osteoblasts and in vitro bone formation from murine embryonic stem cells. Tissue Eng. 7, 89-99.
- Dani, C., Smith, A.G., Dessolin, S., Leroy, P., Staccini, L., Villageois, P., Darimont, C., and Ailhaud, G. (1997). Differentiation of embryonic stem cells into adipocytes in vitro. J. Cell Sci. 110, 1279-1285.
- Dinsmore, J., Ratliff, J., Deacon, T., Pakzaban, P., Jacoby, D., Galpern, W., and Isacson, O. (1996). Embryonic stem cells differentiated in vitro as a novel source of cells for transplantation. Cell Transplant. 5, 131-143.
- Doetsch, F., Caille, I., Lim, D.A., Garcia-Verdugo, J.M., and Alvarez-Buylla, A. (1999). Subventricular zone astrocytes are neural stem cells in the adult mammalian brain. Cell. 97, 703-716.
- Doetschman, T., Eistetter, H., Katz, M., Schmit, W., and Kemler, R. (1985). The *in vitro* development of blastocystderived embryonic stem cell lines: formation of visceral yolk sac, blood islands and myocardium. J. Embryol. Exp. Morph. 87, 27-45.
- Drab, M., Haller, H., Bychkov, R., Erdmann, B., Lindschau, C., Haase, H., Morano, I., Luft, F.C., and Wobus, A.M. (1997). From totipotent embryonic stem cells to spontaneously contracting smooth muscle cells: a retinoic acid and dbcAMP in vitro differentiation model. FASEB J. 11, 905-915.
- Eglitis, M.A. and Mezey, E. (1997). Hematopoietic cells differentiate into both microglia and macroglia in the brains of adult mice. Proc. Natl. Acad. Sci. U. S. A. 94, 4080-4085.
- Erices, A., Conget, P., and Minguell, J.J. (1999).
 Mesenchymal progenitor cells in human umbilical cord blood. Br. J. Haematol. 109, 235-242.
- Evans, M.J. and Kaufman, M.H. (1981). Establishment in culture of pluripotential cells from mouse embryos. Nature. 292, 154-156.
- 24. Fairchild, P.J., Brook, F.A., Gardner, R.L., Graca, L., Strong, V., Tone, Y., Tone, M., Nolan, K.F., and Waldmann, H. (2000). Directed differentiation of dendritic cells from mouse embryonic stem cells. Curr. Biol. 10, 1515-1518.
- Ferrari, G., Cusella-De Angelis, G., Coletta, M., Paolucci, E., Stornaiuolo, A., Cossu, G., and Mavilio, F. (1998). Muscle regeneration by bone marrow-derived myogenic progenitors. Science. 279, 1528-1530.

- Fraichard, A., Chassande, O., Bilbaut, G., Dehay, C., Savatier, P., and Samarut, J. (1995). *In vitro* differentiation of embryonic stem cells into glial cells and functional neurons. J. Cell Sci. 108, 3181-3188.
- Friedenstein, A.J., Gorskaja, U.F., and Kulagina, N.N. (1976).
 Fibroblast precursors in normal and irradiated mouse hematopoietic organs. Exp. Hematol. 4, 267-274.
- Galli, R., Borello, U., Gritti, A., Minasi, M.G., Bjornson, C., Coletta, M., Mora, M., De Angelis, M.G., Fiocco, R., Cossu, G., and Vescovi, A.L. (2000). Skeletal myogenic potential of human and mouse neural stem cells. Nat. Neurosci. 3, 986-991.
- 29. Gottlieb, D.I. and Huettner, J.E. (1999). An *in vitro* pathway from embryonic stem cells to neurons and glia. Cells Tissues Organs. *165*, 165-172.
- Grimaldi, P.A., Teboul, L., Inadera, H., Gaillard, D., and Amri,
 E.Z. (1997). Trans-differentiation of myoblasts to adipoblasts: triggering effects of fatty acids and thiazolidinediones.
 Prostaglandins. Leukot. Essent. Fatty. Acids. 57, 71-75.
- Gussoni, E., Soneoka, Y., Strickland, C.D., Buzney, E.A., Khan, M.K., Flint, A.F., Kunkel, L.M., and Mulligan, R.C. (1999). Dystrophin expression in the mdx mouse restored by stem cell transplantation. Nature. 401, 390-394.
- 32. Hirashima, M., Kataoka, H., Nishikawa, S., Matsuyoshi, N., and Nishikawa, S. (1999). Maturation of embryonic stem cells into endothelial cells in an *in vitro* model of vasculogenesis. Blood. 93, 1253-1263.
- 33. Itskovitz-Eldor, J., Schuldiner, M., Karsenti, D., Eden, A., Yanuka, O., Amit, M., Soreq, H., and Benvenisty, N. (2000). Differentiation of human embryonic stem cells into embryoid bodies comprising the three embryonic germ layers. Mol. Med. 6, 88-95.
- Jackson, K.A., Mi, T., and Goodell, M.A. (1999).
 Hematopoietic potential of stem cells isolated from murine skeletal muscle. Proc. Natl. Acad. Sci. U. S. A. 96, 14482-14486.
- Jackson, K., Majka SM, Wang H, Pocius J, Hartley CJ, Majesky MW, Entman ML, Michael LH, Hirschi KK, and and Goodell MA (2001). Regeneration of ischemic cardiac muscle and vascular endothelium by adult stem cells. J. Clin. Invest. 107, 1-8.
- Johansson, B.M. and Wiles, M.V. (1995). Evidence for involvement of activin A and bone morphogenetic protein 4 in mammalian mesoderm and hematopoietic development. Mol. Cell Biol. 15, 141-151.
- 37. Johansson, C.B., Momma, S., Clarke, D.L., Risling, M., Lendahl, U., and Frisen, J. (1999). Identification of a neural stem cell in the adult mammalian central nervous system. Cell. 96, 25-34.
- Kawasaki, H., Mizuseki, K., Nishikawa, S., Kaneko, S., Kuwana, Y., Nakanishi, S., Nishikawa, S.I., and Sasai, Y. (2000).
 Induction of midbrain dopaminergic neurons from ES cells by stromal cell-derived inducing activity. Neuron. 28, 31-40.

- 39. Kehat, I., Kenyagin-Karsenti, D., Druckmann, M., Segev, H., Amit, M., Gepstein, A., Livne, E., Binah, O., Itskovitz-Eldor, J., and Gepstein, L. (2001). Human embryonic stem cells can differentiate into myocytes portraying cardiomyocytic structural and functional properties. J. Clin. Invest. (in press).
- Kerr, D.A., Llado, J., Shamblott, M., Maragakis, N., Irani, D.N., Dike, S., Sappington, A., Gearhart, J., and Rothstein, J. (2001). Human embryonic germ cell derivatives facillitate motor recovery of rats with diffuse motor neuron injury.
- Klug, M.G., Soonpaa, M.H., Koh, G.Y., and Field, L.J. (1996). Genetically selected cardiomyocytes from differentiating embryonic stem cells form stable intracardiac grafts. J. Clin. Invest. 98, 216-224.
- 42. Kocher, A.A., Schuster, M.D., Szabolcs, M.J., Takuma, S., Burkhoff, D., Wang, J., Homma, S., Edwards, N.M., and Itescu, S. (2001). Neovascularization of ischemic myocardium by human bone-marrow-derived angioblasts prevents cardiomyocyte apoptosis, reduces remodeling and improves cardiac function. Nat. Med. 7, 430-436.
- Kopen, G.C., Prockop, D.J., and Phinney, D.G. (1999).
 Marrow stromal cells migrate throughout forebrain and cerebellum, and they differentiate into astrocytes after injection into neonatal mouse brains. Proc. Natl. Acad. Sci. U. S. A. 96, 10711-10716.
- 44. Kramer, J., Hegert, C., Guan, K., Wobus, A.M., Muller, P.K., and Rohwedel, J. (2000). Embryonic stem cell-derived chondrogenic differentiation *in vitro*: activation by BMP-2 and BMP-4. Mech. Dev. 92, 193-205.
- Krause, D.S., Theise, N.D., Collector, M.I., Henegariu, O., Hwang, S., Gardner, R., Neutzel, S., and Sharkis, S.J. (2001). Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. Cell. 105, 369-377.
- Kuznetsov, S.A., Mankani, M.H., Gronthos, S., Satomura, K., Bianco, P., and Robey P.G. (2001). Circulating skeletal stem cells. J. Cell Biol. 153, 1133-40.
- Lagasse, E., Connors, H., Al Dhalimy, M., Reitsma, M., Dohse, M., Osborne, L., Wang, X., Finegold, M., Weissman, I.L., and Grompe, M. (2000). Purified hematopoietic stem cells can differentiate into hepatocytes *in vivo*. Nat. Med. 6, 1229-1234.
- 48. Lee, S.H., Lumelsky, N., Studer, L., Auerbach, J.M., and McKay, R.D. (2000). Efficient generation of midbrain and hindbrain neurons from mouse embryonic stem cells. Nat. Biotechnol. 18, 675-679.
- 49. Li, M., Pevny, L., Lovell-Badge, R., and Smith, A. (1998). Generation of purified neural precursors from embryonic stem cells by lineage selection. Curr. Biol. 8, 971-974.
- Liechty, K.W., MacKenzie, T.C., Shaaban, A.F., Radu, A., Moseley, A.B., Deans, R., Marshak, D.R., and Flake, A.W. (2000). Human mesenchymal stem cells engraft and demonstrate site-specific differentiation after in utero transplantation in sheep. Nat. Med. 6, 1282-1286.

- 51. Lieschke, G.J. and Dunn, A.R. (1995). Development of functional macrophages from embryonal stem cells *in vitro*. Exp. Hematol. 23, 328-334.
- 52. Liu, S., Qu, Y., Stewart, T.J., Howard, M.J., Chakrabortty, S., Holekamp, T.F., and McDonald, J.W. (2000). Embryonic stem cells differentiate into oligodendrocytes and myelinate in culture and after spinal cord transplantation. Proc. Natl. Acad. Sci. U. S. A. 97, 6126-6131.
- 53. Lumelsky, N., Blondel, O., Laeng, P., Velasco, I., Ravin, R., and McKay, R. (2001). Differentiation of Embryonic Stem Cells to Insulin-Secreting Structures Similiar to Pancreatic Islets. Science. 292, 1389-1394.
- 54. Makino, S., Fukuda, K., Miyoshi, S., Konishi, F., Kodama, H., Pan, J., Sano, M., Takahashi, T., Hori, S., Abe, H., Hata, J., Umezawa, A., and Ogawa, S. (1999). Cardiomyocytes can be generated from marrow stromal cells in vitro. J. Clin. Invest. 103, 697-705.
- 55. Maltsev, V.A., Rohwedel, J., Hescheler, J., and Wobus, A.M. (1993). Embryonic stem cells differentiate *in vitro* into cardiomyocytes representing sinusnodal, atrial and ventricular cell types. Mech. Dev. 44, 41-50.
- Matsui, Y., Zsebo, K., and Hogan, B.L. (1992). Derivation of pluripotential embryonic stem cells from murine primordial germ cells in culture. Cell. 70, 841-847.
- McBurney, M.W., Reuhl, K.R., Ally, A.I., Nasipuri, S., Bell, J.C., and Craig, J. (1988). Differentiation and maturation of embryonal carcinoma-derived neurons in cell culture. J. Neurosci. 8, 1063-1073.
- McCune, J.M., Namikawa, R., Kaneshima, H., Shultz, L.D., Lieberman, M., and Weissman, I.L. (1988). The SCID-hu mouse: murine model for the analysis of human hematolymphoid differentiation and function. Science. 241, 1632-1639.
- Megeney, L.A., Kablar, B., Garrett, K., Anderson, J.E., and Rudnicki, M.A. (1996). MyoD is required for myogenic stem cell function in adult skeletal muscle. Genes Dev. 10, 1173-1183.
- Mezey, E., Chandross, K.J., Harta, G., Maki, R.A., and McKercher, S.R. (2000). Turning blood into brain: cells bearing neuronal antigens generated *in vivo* from bone marrow. Science. 290, 1779-1782.
- 61. Morrison, S.J., Uchida, N., and Weissman, I.L. (1995). The biology of hematopoietic stem cells. Annu. Rev. Cell. Dev. Biol. 11, 35-71.
- 62 Namikawa, R., Weilbaecher, K.N., Kaneshima, H., Yee, E.J., and McCune, J.M. (1990). Long-term human hematopoiesis in the SCID-hu mouse. J. Exp. Med. *172*, 1055-1063.
- 63. O'Shea, K.S. (1999). Embryonic stem cell models of development. Anat. Rec. 257, 32-41.
- 64. Orlic, D., Kajstura, J., Chimenti, S., Jakoniuk, I., Anderson, S.M., Li, B., Pickel, J., McKay, R., Nadal-Ginard, B., Bodine, D.M., Leri, A., and Anversa, P. (2001). Bone marrow cells regenerate infarcted myocardium. Nature. 410, 701-705.

- 65. Palmer, T.D., Schwartz, P.H., Taupin, P., Kaspar, B., Stein, S.A., and Gage, F.H. (2001). Progenitor cells from human brain after death. Nature. *411*, 42-43.
- Pera, M.F., Cooper, S., Mills, J., and Parrington, J.M. (1989).
 Isolation and characterization of a multipotent clone of human embryonal carcinoma cells. Differentiation. 42, 10-23.
- Pera, M.F. and Herszfeld, D. (1998). Differentiation of human pluripotent teratocarcinoma stem cells induced by bone morphogenetic protein-2. Reprod. Fertil. Dev. 10, 551-555.
- 68. Pereira, R.F., Halford, K.W., O'Hara, M.D., Leeper, D.B., Sokolov, B.P., Pollard, M.D., Bagasra, O., and Prockop, D.J. (1995). Cultured adherent cells from marrow can serve as long-lasting precursor cells for bone, cartilage, and lung in irradiated mice. Proc. Natl. Acad. Sci. U. S. A. 92, 4857-4861.
- 69. Perkins, A.C. (1998). Enrichment of blood from embryonic stem cells *in vitro*. Reprod. Fertil. Dev. 10, 563-572.
- Petersen, B.E., Bowen, W.C., Patrene, K.D., Mars, W.M., Sullivan, A.K., Murase, N., Boggs, S.S., Greenberger, J.S., and Goff, J.P. (1999). Bone marrow as a potential source of hepatic oval cells. Science. 284, 1168-1170.
- Pittenger, M.F., Mackay, A.M., Beck, S.C., Jaiswal, R.K., Douglas, R., Mosca, J.D., Moorman, M.A., Simonetti, D.W., Craig, S., and Marshak, D.R. (1999). Multilineage potential of adult human mesenchymal stem cells. Science. 284, 143-147.
- 72. Potocnik, A.J., Nielsen, P.J., and Eichmann, K. (1994). *In vitro* generation of lymphoid precursors from embryonic stem cells. EMBO J. *13*, 5274-5283.
- Prelle, K., Wobus, A.M., Krebs, O., Blum, W.F., and Wolf, E. (2000). Overexpression of insulin-like growth factor-II in mouse embryonic stem cells promotes myogenic differentiation. Biochem. Biophys. Res. Commun. 277, 631-638.
- Prockop, D.J. (1997). Marrow stromal cells as stem cells for nonhematopoietic tissues. Science. 276, 71-74.
- Ramiya, V.K., Maraist, M., Arfors, K.E., Schatz, D.A., Peck, A.B., and Cornelius, J.G. (2000). Reversal of insulin-dependent diabetes using islets generated in vitro from pancreatic stem cells. Nat. Med. 6, 278-282.
- Rathjen, P.D., Lake, J., Whyatt, L.M., Bettess, M.D., and Rathjen, J. (1998). Properties and uses of embryonic stem cells: prospects for application to human biology and gene therapy. Reprod. Fertil. Dev. 10, 31-47.
- Reubinoff, B.E., Pera, M.F., Fong, C.Y., Trounson, A., and Bongso, A. (2000). Embryonic stem cell lines from human blastocysts: somatic differentiation in vitro. Nat. Biotechnol. 18, 399-404.
- Reynolds, B.A. and Weiss, S. (1996). Clonal and population analyses demonstrate that an EGF-responsive mammalian embryonic CNS precursor is a stem cell. Dev. Biol. 175, 1-13.

- Reynolds, J.N., Ryan, P.J., Prasad, A., and Paterno, G.D. (1994). Neurons derived from embryonal carcinoma (P19) cells express multiple GABA(A) receptor subunits and fully functional GABA(A) receptors. Neurosci. Lett. 165, 129-132.
- Risau, W., Sariola, H., Zerwes, H.G., Sasse, J., Ekblom, P., Kemler, R., and Doetschman, T. (1988). Vasculogenesis and angiogenesis in embryonic-stem-cell-derived embryoid bodies. Development. 102, 471-478.
- 81. Roach, S., Schmid, W., and Pera, M.F. (1994). Hepatocytic transcription factor expression in human embryonal carcinoma and yolk sac carcinoma cell lines: expression of HNF-3α in models of early endodermal cell differentiation. Exp. Cell. Res. 215, 189-198.
- Rohwedel, J., Maltsev, V., Bober, E., Arnold, H.H., Hescheler, J., and Wobus, A.M. (1994). Muscle cell differentiation of embryonic stem cells reflects myogenesis in vivo: developmentally regulated expression of myogenic determination genes and functional expression of ionic currents. Dev. Biol. 164, 87-101.
- 83. Sanchez-Ramos, J., Song, S., Cardozo-Pelaez, F., Hazzi, C., Stedeford, T., Willing, A., Freeman, T.B., Saporta, S., Janssen, W., Patel, N., Cooper, D.R., and Sanberg, P.R. (2000). Adult bone marrow stromal cells differentiate into neural cells in vitro. Exp. Neurol. 164, 247-256.
- 84. Schuldiner, M., Yanuka, O., Itskovitz-Eldor, J., Melton, D., and Benvenisty, N. (2000). Effects of eight growth factors on the differentiation of cells derived from human embryonic stem cells. Proc. Natl. Acad. Sci. U. S. A. 97, 11307-11312.
- 85. Seale, P., Sabourin, L.A., Girgis-Gabardo, A., Mansouri, A., Gruss, P., and Rudnicki, M.A. (2000). Pax7 is required for the specification of myogenic satellite cells. Cell. 102, 777-786.
- Shamblott, M.J., Axelman, J., Littlefield, J.W., Blumenthal, P.D., Huggins, G.R., Cui, Y., Cheng, L., and Gearhart, J.D. (2001). Human embryonic germ cell derivatives express a broad range of develpmentally distinct markers and proliferate extensively *in vitro*. Proc. Natl. Acad. Sci. U. S. A. 98, 13-118.
- Shamblott, M.J., Axelman, J., Wang, S., Bugg, E.M., Littlefield, J.W., Donovan, P.J., Blumenthal, P.D., Huggins, G.R., and Gearhart, J.D. (1998). Derivation of pluripotent stem cells from cultured human primordial germ cells. Proc. Natl. Acad. Sci. U. S. A. 95, 13726-13731.
- 88. Slager, H.G., Van Inzen, W., Freund, E., Van den Eijnden-Van Raaij A.J.M., and Mummery, C.L. (1993). Transforming growth factor-beta in the early mouse embryo: implications for the regulation of muscle formation and implantation. Dev. Genet. 14, 212-224.
- 89. Soria, B., Roche, E., Berna, G., Leon-Quinto, T., Reig, J.A., and Martin, F. (2000). Insulin-secreting cells derived from embryonic stem cells normalize glycemia in streptozotocin-induced diabetic mice. Diabetes. 49, 157-162.
- Spangrude, G.J., Smith, L., Uchida, N., Ikuta, K., Heimfeld, S., Friedman, J., and Weissman, I.L. (1991). Mouse hematopoietic stem cells. Blood. 78, 1395-1402.

- 91. Strubing, C., Ahnert-Hilger, G., Shan, J., Wiedenmann, B., Hescheler, J., and Wobus, A.M. (1995). Differentiation of pluripotent embryonic stem cells into the neuronal lineage *in vitro* gives rise to mature inhibitory and excitatory neurons. Mech. Dev. 53, 275-287.
- Taniguchi, H., Toyoshima, T., Fukao, K., and Nakauchi, H. (1996). Presence of hematopoietic stem cells in the adult liver. Nat. Med. 2, 198-203.
- 93. Theise, N.D., Nimmakayalu, M., Gardner, R., Illei, P.B., Morgan, G., Teperman, L., Henegariu, O., and Krause, D.S. (2000). Liver from bone marrow in humans. Hepatology. 32, 11-16.
- Thompson, S., Stern, P.L., Webb, M., Walsh, F.S., Engstrom, W., Evans, E.P., Shi, W.K., Hopkins, B., and Graham, C.F. (1984). Cloned human teratoma cells differentiate into neuron-like cells and other cell types in retinoic acid. J. Cell. Sci. 72, 37-64.
- Thomson, J.A., Itskovitz-Eldor, J., Shapiro, S.S., Waknitz, M.A., Swiergiel, J.J., Marshall, V.S., and Jones, J.M. (1998).
 Embryonic stem cell lines derived from human blastocysts. Science. 282, 1145-1147.
- Tomita, S., Li, R.K., Weisel, R.D., Mickle, D.A., Kim, E.J., Sakai, T., and Jia, Z.Q. (1999). Autologous transplantation of bone marrow cells improves damaged heart function 672. Circulation. 100 (Suppl. II), 11247-11256.
- Trojanowski, J.Q., Mantione, J.R., Lee, J.H., Seid, D.P., You, T., Inge, L.J., and Lee, V.M. (1993). Neurons derived from a human teratocarcinoma cell line establish molecular and structural polarity following transplantation into the rodent brain. Exp. Neurol. 122, 283-294.
- 98. Tsai, M., Wedemeyer, J., Ganiatsas, S., Tam, S.Y., Zon, L.I., and Galli, S.J. (2000). *In vivo* immunological function of mast cells derived from embryonic stem cells: an approach for the rapid analysis of even embryonic lethal mutations in adult mice *in vivo*. Proc. Natl. Acad. Sci. U. S. A. 97, 9186-9190.
- Uchida, N., Buck, D.W., He, D., Reitsma, M.J., Masek, M., Phan, T.V., Tsukamoto, A.S., Gage, F.H., and Weissman, I.L. (2000). Direct isolation of human central nervous system stem cells. Proc. Natl. Acad. Sci. U. S. A. 97, 14720-14725.
- 100. Wakitani, S., Saito, T., and Caplan, A.I. (1995). Myogenic cells derived from rat bone marrow mesenchymal stem cells exposed to 5-azacytidine 754. Muscle. Nerve. 18, 1417-1426.
- 101. Wang, X., Al-Dhalimy, M., Lagasse, E., Finegold, M., and Grompe, M. (2001). Liver repopulation and correction of metabolic liver disease by transplanted adult mouse pancreatic cells. Am. J. Pathol. 158, 571-579.
- 102. Weiss, S., Dunne, C., Hewson, J., Wohl, C., Wheatley, M., Peterson, A.C., and Reynolds, B.A. (1996). Multipotent CNS stem cells are present in the adult mammalian spinal cord and ventricular neuroaxis. J. Neurosci. 16, 7599-7609.

- 103. Westfall, M.V., Pasyk, K.A., Yule, D.I., Samuelson, L.C., and Metzger, J.M. (1997). Ultrastructure and cell-cell coupling of cardiac myocytes differentiating in embryonic stem cell cultures. Cell. Motil. Cytoskeleton. 36, 43-54.
- 104. Wiles, M.V. and Keller, G. (1991). Multiple hematopoietic lineages develop from embryonic stem (ES) cells in culture. Development. *111*, 259-267.
- 105. Wobus, A.M., Rohwedel, J., Maltsev, V., and Hescheler, J. (1995). Development of cardiomyocytes expressing cardiac-specific genes, action potentials, and ionic channels during embryonic stem cell-derived cardiogenesis. Ann. N. Y. Acad. Sci. 752, 460-469.
- Woodbury, D., Schwarz, E.J., Prockop, D.J., and Black, I.B. (2000). Adult rat and human bone marrow stromal cells differentiate into neurons. J. Neurosci. Res. 61, 364-370.
- Yamane, T., Hayashi, H., Mizoguchi, M., Yamazaki, H., and Kunisada, T. (1999). Derivation of melanocytes from embryonic stem cells in culture. Dev. Dyn. 216, 450-458.

- 108. Yamashita, J., Itoh, H., Hirashima, M., Ogawa, M., Nishikawa, S., Yurugi, T., Naito, M., Nakao, K., and Nishikawa, S. (2000). Flk1-positive cells derived from embryonic stem cells serve as vascular progenitors. Nature. 408, 92-96.
- 109. Zuk, P.A., Zhu, M., Mizuno, H., Huang, J., Futrell, J.W., Katz, A.J., Benhaim, P., Lorenz, H.P., and Hedrick, M.H. (2001). Multilineage cells from human adipose tissue: implications for cell-based therapies. Tissue Eng. 7, 211-228.
- 110. Zulewski, H., Abraham, E.J., Gerlach, M.J., Daniel, P.B., Moritz, W., Muller, B., Vallejo, M., Thomas, M.K., and Habener, J.F. (2001). Multipotential nestin-positive stem cells isolated from adult pancreatic islets differentiate ex vivo into pancreatic endocrine, exocrine, and hepatic phenotypes. Diabetes. 50, 521-533.